

Virtual Screening and Modeling of Phosphoglycerate Mutase 1

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Abstract Methods Phosphoglycerate Mutase 1 (PGAM1) is a glycolytic protein Docking with Smina upregulated in many types of cancer cells. In a phenomenon imulatior called the Warburg Effect, cancer cells have a higher rate of glycolysis than healthy cells. Altering PGAM1 activity can change cancer cell proliferation and thus is an important pharmacological target for cancer suppression. A small Steered MD molecule, MJE3, inhibits PGAM1 activity and decreases cancer cell proliferation. A binding model of MJE3 to PGAM1 was created using molecular dynamic simulations and protein-PGAM1 Crystallized Structure – 4GPZ ligand dockings. Key interactions were used to create pharmacophore models and perform virtual screenings of purchasable compounds for potential new drug targets of PGAM1 inhibition.

Introduction/Background

PGAM1 converts 3PG to 2PG during glycolysis. 3PG and 2PG are important substrates for metabolic and biosynthetic pathways. Inhibition of PGAM1 increases levels of 3PG and decreases levels of 2PG resulting in inhibition of 6PGD (pentose phosphate pathway). Decreased activity in this metabolic pathway decreases levels of biosynthetic materials important for cancer cells to be able to rapidly divide.



Unbound State of MJE3 (Sorensen and Cravatt)

Covalently Bound State of MJE3 to Lysine 100

MJE3 inhibits PGAM1 by covalently binding to lysine 100 on PGAM1 (circled in yellow). The epoxide bond on MJE3 is broken open by the nitrogen on lysine 100. MJE3 is hydrolyzed in situ of its benzyl ester substituent before binding (circled in blue). Currently there are no crystallized structures of MJE3 bound to

phosphate pathway

Role of PGAM1 in metabolic pathways

(Li and Ren)





PGAM1. A complete model will give us more insight to MJE3's action and lead us to find other drug alternatives. Sorensen and Cravatt have created a docking model, but have not used it to search for other binding agents. We aim to create a model of our own to compare to the Sorensen and Cravatt model while also finding drugs that dock similarly.

We docked MJE3 to PGAM1 to sample conformations and poses that will produce models of binding. Docking software holds the receptor rigid, so molecular dynamics simulations are useful to move the receptor and ligand into more energy efficient and realistic poses. Given a structural model, we performed a virtual screen of available compounds using pharmacophore models. A pharmacophore is a group of features of a compound that show the interactions between other molecules such as hydrogen binding and hydrophobes.

Analogues of MJE3 with alterations in the indole side chain do not effectively label PGAM1. Our docking and pharmacophore models will focus around the epoxide and indole groups, for we know that those are important groups for binding.

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R117 of PGAM1. Our top 3 dockings also had the indole group docked near those amino acids, but flipped to different orientations. In our models, the hydrogen on the nitrogen of the indole group made hydrogen bonds with double bonded oxygen on the backbone.

Results – Selected Compounds









7 compounds were selected based on our hypothesis of MJE3 binding

- Hydrogen accepting group at same location as site of attack by Lysine 100
- Hydrogen donating and hydrophobic or aromatic region where indole group of MJE3 is docked

Other criteria for compounds

- Less than -6 minimized affinity
- Less than 2 minimized RMSD
- Favorable hydrogen bonds throughout the compound
- Less than 500 g/mol Molar Mass

Compound Number	N	linimized Affinity	Minimized RMSD
	1	-8.27346	1.92313
	2	-7.97418	1.77765
	3	-7.11071	1.54039

Future Directions

The selected compounds will be purchased from MolPort and tested for PGAM1 inhibition.

References

Evans, M. J., Morris, G. M., Wu, J., Olson, A. J., Sorensen, E. J., & Cravatt, B. F. (2007). Mechanistic and structural requirements for active site labeling of phosphoglycerate mutase by spiroepoxides. *Molecular bioSystems, 7,* 495–506.

Hitosugi, T., Zhou, L., Elf, S., Fan, J., Kang, H. B., Seo, J. H., Shan, C., Dai, Q., Zhang, L., Xie, J., Gu, T. L., Jin, P., Aleä koviä‡, M., LeRoy, G., Kang, Y., Sudderth, J. A., DeBerardinis, R. J., Luan, C. H., Chen, G. Z., Muller, S., Shin, D. M., Owonikoko, T. K., Lonial, S., Arellano, M. L., Khoury, H. J., Khuri, F. R., Lee, B. H., Ye, K., Boggon, T. J., Kang, S., He, C., & Chen, J. (2012). Phosphoglycerate mutase 1 coordinates glycolysis and biosynthesis to promote tumor growth. Cancer *cell, 5,* 585–600.

Hitosugi, T., Zhou, L., Fan, J., Elf, S., Zhang, L., Xie, J., Wang, Y., Gu, T. L., Aleä koviä[‡], M., LeRoy, G., Kang, Y., Kang, H. B., Seo, J. H., Shan, C., Jin, P., Gong, W., Lonial, S., Arellano, M. L., Khoury, H. J., Chen, G. Z., Shin, D. M., Khuri, F. R., Boggon, T. J., Kang, S., He, C., & Chen, J. (2013). Tyr26 phosphorylation of PGAM1 provides a metabolic advantage to tumours by stabilizing the active conformation. *Nature* communications, , 1790.

Jiang, X., Sun, Q., Li, H., Li, K., & Ren, X. (2013). The

role of phosphoglycerate mutase 1 in tumor aerobic glycolysis and its potential therapeutic implications. International journal of cancer. Journal international du cancer, , .